XerD unloads bacterial SMC complexes at the replication terminus

Graphical Abstract

Highlights
- *Bacillus subtilis* XerD binds novel sites in the replication terminus region
- SMC complexes loaded at the origin are unloaded by XerD at the terminus
- XerD functions as a site-specific unloader of translocating SMC complexes
- ParB-mediated condensin loading and XerD-mediated condensin unloading are conserved

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In Brief
SMC condensin complexes are loaded at replication origins by the partitioning protein ParB. Karaboja et al. show that condensins are unloaded when they reach the terminus by the recombinase XerD. Thus, broadly conserved factors that act at the origin and terminus load and unload SMC complexes that travel between them.
XerD unloads bacterial SMC complexes at the replication terminus

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SUMMARY

Bacillus subtilis structural maintenance of chromosomes (SMC) complexes are topologically loaded at centromeric sites adjacent to the replication origin by the partitioning protein ParB. These ring-shaped ATPases then translocate down the left and right chromosome arms while tethering them together. Here, we show that the site-specific recombinase XerD, which resolves chromosome dimers, is required to unload SMC tethers when they reach the terminus. We identify XerD-specific binding sites in the terminus region and show that they dictate the site of unloading in a manner that depends on XerD but not its catalytic residue, its partner protein XerC, or the recombination site dif. Finally, we provide evidence that ParB and XerD homologs perform similar functions in Staphylococcus aureus. Thus, two broadly conserved factors that act at the origin and terminus have second functions in loading and unloading SMC complexes that travel between them.

INTRODUCTION

Structural maintenance of chromosomes (SMC) complexes play central roles in organizing genomes in all domains of life (Hirano, 2016; Uhlmann, 2016; Yatskevich et al., 2019). Recent studies in bacteria and eukaryotes have provided evidence for a model in which these ring-shaped ATPases structure chromosomes by extruding DNA loops (Hassler et al., 2018; Marko et al., 2019; Yatskevich et al., 2019). In this model, SMC complexes are topologically loaded onto DNA and then translocate away from their loading site, processively enlarging a chromosome loop. Loop extrusion provides an explanation of how these complexes resolve newly replicated origins in bacteria, linearly compact and resolve sister chromatids in eukaryotic prophase, and generate topological associating domains (TADs) during interphase. Although factors that load SMC complexes onto DNA at specific sites along the chromosome have been well characterized (Yatskevich et al., 2019), site-specific unloaders that could specify domain boundaries have only been recently hinted at (Lioy et al., 2018; Mäkelä and Sherratt, 2020; Nolivos et al., 2016). Here, we report that the XerD recombinase is required for site-specific unloading of bacterial SMC complexes at the replication terminus.

In most bacteria, a small set of broadly conserved factors act at the origin and terminus to promote the resolution and segregation of replicated chromosomes (Badrinarayanan et al., 2015; Reyes-Lamothe et al., 2012). The ParAB/parS system plays a central role in segregating newly replicated origins. ParB binds site-specifically to centromeric sequences called parS adjacent to the replication origin, and ParA ATPases act on these complexes, promoting their directed movement toward opposite cell poles. The XerCD/dif system functions at the terminus to resolve fully replicated chromosomes (Lesterlin et al., 2004; Mondonet and Barre, 2014). XerC and XerD have been extensively studied in E. coli, although some of the findings have been reproduced in B. subtilis and other organisms (Blakely et al., 1993; Cornet et al., 1997; Sciochetti et al., 1999, 2001; Val et al., 2008). XerC and XerD are site-specific recombinases that bind a recombination site called dif and catalyze the resolution of chromosome dimers that arise as a result of homologous recombination during replication (Blakely et al., 1993; Cornet et al., 1997; Sciochetti et al., 1999, 2001). The dif site is located in the replication terminus region, as defined by the sites bound by the replication termination protein RTP (Duggin et al., 2008), which is segregated by the SpoIIIIE/FtsK family of proteins at the time of cell division (Crozat et al., 2014; Sherratt et al., 2010). Repeated rounds of XerCD-mediated recombination at dif have also been shown to remove interlinks (known as catenanes) between sister chromosomes (Grainge et al., 2007; Ip et al., 2003; Shimokawa et al., 2013). Studies in a growing number of bacteria have revealed that the ParB/parS nucleoprotein complex performs a second function: the recruitment and loading of the bacterial SMC complex (Bohm et al., 2020; Gruber and Errington, 2009; Minnen et al., 2011; Sullivan et al., 2009;
Tran et al., 2017). Once loaded, SMC complexes travel from origin-proximal parS sites to the terminus while tethering the left and right chromosome arms together (Wang et al., 2015, 2017). Thus, these complexes generate a single chromosome loop that draws sister chromosomes in on themselves and away from each other and likely facilitate the removal of pre-catenanes by Topoisomerase IV (Burmann and Gruber, 2015; Marko, 2009). The fate of SMC complexes upon arrival at the terminus has been unclear.

Chromatin immunoprecipitation sequencing (ChIP-seq) and Hi-C studies in B. subtilis suggest that loop extrusion by SMC complexes is mediated by two “handcuffed” rings (Branda˜o et al., 2019; Wang et al., 2015). Biochemical studies indicate that the E. coli MukBEF complex, an analog of SMC complexes, indeed forms dimers (Badrinarayanan et al., 2012; Rajasekar et al., 2019). In this model, separate SMC complexes encircle the DNA on either side of parS and translocate away from the loading site as a dimer. If correct, the tethered rings would remain topologically associated with the DNA when they reach the terminus and might require an active mechanism to unload them. Here, we show that the XerD recombinase has a heretofore unrecognized second function in unloading bacterial SMC complexes at the terminus. We identify XerD-specific binding sites in the terminus region and show that these binding sites and XerD but not XerC or dif are required for site-specific unloading of SMC complexes. We identify similar binding sites in S. aureus and provide evidence that ParB and XerD have similar loading and unloading functions in this bacterium. Thus, the loading and unloading of SMC complexes are mediated by two broadly conserved factors: one involved in origin segregation, the other in terminus resolution. Similar site-specific unloading activities could modulate eukaryotic SMC complexes during interphase and mitosis.

RESULTS

B. subtilis cells lacking their endogenous parS sites and harboring a single ectopic parS site juxtapose large tracks of DNA flanking the ectopic site in a ParB- and SMC-dependent manner (Wang et al., 2015, 2017) (Figures 1A and S1A). Intriguingly, the DNA juxtaposition never extends beyond a ~300 kb region surrounding the replication terminus, regardless of where parS is inserted (Figure 1A). Similarly, SMC complexes are specifically enriched along the juxtaposed DNA and enrichment does not extend beyond the terminus region (Wang et al., 2017) (Figures 1B and S1B).

To investigate whether any of the factors that act in the terminus region also function to prevent SMC from translocating...
beyond it, we analyzed SMC enrichment in a set of mutants harboring a single parS site at \(-117^\circ\). We performed ChIP-seq on cells lacking the replication termination protein RTP, which binds Ter sites and ensures replication terminates within a defined region (Duggin et al., 2008); the two DNA translocases (SpoIIIE and SftA) (Kaimer et al., 2011) that act at the division septum to segregate terminus-proximal DNA into the appropriate daughter cell; and the site-specific recombinases XerC (CocV) and XerD (RipX) and the recombination site dif (Sciochetti et al., 1999, 2001). SMC enrichment was largely unaffected in cells lacking RTP or the two DNA translocases (Figures 1B and S1C). In cells lacking dif or XerC, SMC enrichment was reduced in regions near parS, but the extent of enrichment was largely unchanged (Figure S1C). SMC enrichment in the \(\Delta xerD\) mutant was even more reduced near parS and appeared to modestly extend beyond the terminus region (Figure 1B, black caret). DNA juxtaposition, as assayed by Hi-C, was also altered in the \(\Delta xerD\) mutant, proceeding modestly but reproducibly beyond the terminus region (Figure S1D). Furthermore, we observed a reduction in DNA juxtaposition in the terminus region in wild-type cells that was largely absent in the \(\Delta xerD\) mutant (Figure S1E). These results prompted us to explore a potential role for XerC/Dif in preventing SMC rings from traveling beyond the terminus. 

Consistent with previous in vitro studies (Blakely et al., 1993; Sciochetti et al., 2001), ChIP-seq with a XerC-GFP fusion strain identified a single XerC enrichment peak centered at dif (Figure 2A) that was dependent on XerD and dif (Figures 2A and S2C). In contrast, a XerD-GFP fusion not only had an enrichment peak at dif but also had five additional peaks in the terminus region (Figure 2B). Similar results were obtained with XerC and XerD fusions to a Protein C (PrC) epitope (Griffin et al., 1981; Zheng et al., 2018)(Figures S2A and S2B). Although XerD enrichment at dif depended on XerC and dif, enrichment at the other five sites was independent of these factors (Figures 2B and S2D). Four of these XerD-specific enrichment peaks spanned a 316 kb region and were present in all ChIP-seq experiments, while the fifth was variable and always weaker than the other four (Figures 2B, 2C, and S2B). Statistical analysis indicates that XerD’s enrichment at \(\Delta xDS1\)–4 sites was significant in all three replicates; enrichment at \(\Delta xDS1\) was significant in two of three replicates (Figure S2E). Motif discovery algorithms from the MEME suite (http://meme-suite.org/) identified a putative binding site (named XDS, for XerD binding site) that was present within each enrichment peak and absent from the rest of the genome (Figure 2D). These XDS sites contain the XerD binding motif at dif adjacent to a sequence that does not resemble the XerC binding motif (Figure 2D). Deleting the XDS1 sequence resulted in the loss of the XerD enrichment peak at 1,717 kb, and inserting XDS1 at 2,540 kb generated a new peak at this ectopic position (Figure 2C).

The XerD-XDS interaction was further analyzed in \(E.\ coli\) via a reporter \(P_{\Delta xDS1-lacZ}\) in which XDS1 was inserted between the -10 and -35 elements of a synthetic promoter (Cho and Bernhardt, 2013) such that transcription of lacZ is repressed if a protein binds to XDS1 (Figure 2E). \(B.\ subtilis\) XerD was expressed under IPTG control, and LacZ production was analyzed on LB agar plates containing X-gal. In the absence of IPTG, the \(E.\ coli\) colonies were blue (Figure 2E); in its presence, the colonies were white. In contrast, expression of \(E.\ coli\) XerD or \(B.\ subtilis\) XerD using identical promoter fusions failed to repress transcription (Figure 2E). We conclude that XerD binds directly to the XDS sites and appears to do so without additional co-factors.

\(B.\ subtilis\) cells lacking XerC or dif grow similarly to the wild-type (Figure 3A), and their chromosomes (called nucleoids) have relatively homogeneous morphologies (Sciochetti et al., 1999) (Figures 3B, 3C, S2G, and S3). In contrast, cells lacking XerD are growth defective, and their nucleoids are heterogeneous in size and appear distended (Lemon et al., 2001; Sciochetti et al., 1999) (Figures 3A–3C, S2G, and S3). Importantly, the growth defect of \(\Delta xerD\) was not suppressed by deletions of xerC (Cornet et al., 1997) (Figure S2H) or recA (Blakely et al., 1993; Kuempel et al., 1991) (Figure S2I), consistent with the idea that XerD has a second function outside of dimer resolution. To investigate whether these phenotypic differences relate to XerD bound to XDS sites, we analyzed a strain in which all five sites were deleted (\(\Delta xDS45\)). The growth and nucleoid morphologies of the mutant cells were similar to those lacking XerC or dif (Figures 3A–3C, S2G, and S3). However, when the XDS deletions were combined with \(\Delta xerC\), \(\Delta dif\), or \(\Delta xerD\), the cells largely resembled the \(\Delta xerD\) mutant (Figures 3A–3C, S2G, and S3). Analysis of the replication origins revealed that the distended nucleoids in the \(\Delta xerD\) single mutant and the double mutants that lack the XDS sites contained multiple origins (Figure 3D). As the DNA replication profiles of these strains were similar to wild-type (Figure S2F), these findings indicate that chromosome segregation is impaired in the mutants. Thus, in addition to its role in dimer resolution, XerD has a second function in chromosome segregation that requires its XerD-specific binding sites.

Close examination of the SMC ChIP-seq profile in the strain harboring the \(-117^\circ\) parS suggests that XerD’s second function is related to condensin. Five statistically significant SMC enrichment peaks in the terminus region coincided with XerD enrichment peaks at dif and the four strong XDS sites (Figures 4A and S4A–S4C). At higher resolution, the XerD and SMC enrichment peaks were superimposable (Figures 4C and S4D). Furthermore, SMC enrichment at the XDS sites varied between XerD and XDS (Figures 4A–4C, S4D, and S4E), while enrichment at dif required XerC, XerD, and dif (Figures 4A and S4E). Importantly, all five SMC enrichment peaks were also present in wild-type and a strain harboring ectopic parS sites at \(-153^\circ\) or \(-94^\circ\) (Figure S4A). In contrast, there was no correlation between SMC enrichment in the terminus region and the Ter sites bound by RTP (Figure S4B), and in strains lacking RTP or the DNA translocases SpoIIIE and SftA, the SMC enrichment peaks at XDS sites were intact (Figure 4B). Finally, the overall ChIP-seq profile in the strain lacking all five XDS sites and harboring a parS site at \(-117^\circ\) was similar to the profile in the \(\Delta xerD\) mutant (Figures 4A and 4B). Altogether, these data suggest that SMC complexes are acted upon by XerD bound to XDS in the terminus region.

XerD, in complex with XerC, not only resolves chromosome dimers at dif sites but has also been found to help resolve catenated chromosomes in \(E.\ coli\) (Grainge et al., 2007; Ip et al., 2003; Shimokawa et al., 2013). Inefficient or impaired removal of these topological barriers could explain why DNA juxtaposition extended so modestly beyond the terminus region in the \(\Delta xerD\) mutant (Figure S1D, black carets). To circumvent potential topological barriers, we used a high-resolution DNA molecule to analyze DNA juxtaposition in the terminus region.
impediments to SMC movement, we investigated whether recruitment of XerD to an ectopic position blocks DNA juxtaposition and SMC translocation. Arrays of XDS sites were inserted at /C0/C109/C14 in a strain with a single parS site at /C0/C94/C14, 175 kb away. DNA interaction frequencies from the /C0/C94 parS to the /C0/C109/C14 XDS array (and its juxtaposed locus at /C0/C84/C14) were similar to those in a strain without an array (Figures 5A–5C, S5A, and S5B). However, interactions beyond these positions were reduced to ~50% with an array of 4 XDS sites and to background level with an array of 12 sites (Figures 5B, 5C, S5A, and S5B). We note that the XDS arrays also generated a chromosome interaction boundary that was SMC and XerD dependent (Figures S5C and S5F), suggesting a distinct role for SMC complexes in short-range interactions along the chromosome (Lioy et al., 2018). SMC enrichment as assayed by ChIP-seq was largely restricted to the region between /C0/C84/C14 and /C0/C109/C14 (Figures 5B and 5C). Importantly, the block to DNA juxtaposition and SMC enrichment required XerD but not XerC (Figure S5F). Similar results were obtained when the (XDS)12 array was inserted at /C0/C80/C14 (Figure S6A) or in a strain that contained a parS site at /C0/C59/C14 and an (XDS)12 array at /C0/C109/C14 (Figures S6B and S6C). An even more dramatic loss of DNA juxtaposition was observed when the (XDS)12 array was inserted at +26/C14 or /C0/C19/C14 in a strain harboring a native parS site at /C0/C1/C14 (Figures 5E and S5D). Importantly, DNA juxtaposition and SMC enrichment were not affected by TetR-CFP fusions bound to an array of 48 tetO operators (Figure S5E) and marker frequency analysis indicates that the (XDS)12 array did not impair DNA replication (Figure S5G). We note that strains with one or two ectopic XDS sites did not impair SMC enrichment (Figure S6D), perhaps because of the large step size predicted for these translocases (see Discussion) (Davidson et al., 2019; Ganji et al., 2018; Kim et al., 2019; Terakawa et al., 2017).

Figure 2. Identification of XerD-specific binding sites at replication terminus
(A) Enrichment profiles from anti-GFP ChIP-seq performed in strains containing xerC-gfp in otherwise wild-type (WT) (top), Δdif (middle), or ΔxerD (bottom) strains. Sequencing reads from ChIP and input samples were normalized to the total number of reads. The ChIP enrichment (ChIP/Input) is shown in 1 kb bins. The position of the dif site is indicated by a red arrow.
(B) Enrichment profiles from anti-GFP ChIP-seq performed in strains containing xerD-gfp in otherwise WT (top), Δdif (middle), or ΔxerC (bottom) strains. XerD-GFP is enriched at four sites (XDS1–4) in addition to dif. A fifth XerD binding site (XDS5) is present in a subset of experiments (B, bottom; C, bottom; Figure S2B). XDS sites do not appear to have a specific orientation. XDS1 (+strand), dif (+strand), XDS2 (−strand), XDS3 (+strand), XDS4 (−strand), XDS5 (−strand). Quantitative analysis of ChIP-seq experiments can be found in Figure S2E.
(C) Enrichment profiles from anti-GFP ChIP-seq plots in strains containing xerD-gfp in otherwise WT (top, biological replicate to the top panel of B), a strain with XDS1 deleted (middle), and a strain with XDS1 inserted at 2,540 kb (bottom).
(D) Top: logo of XerD binding site generated with XDS1–4. Bottom: the five XerD binding sites compared with B. subtilis dif with indicated XerC and XerD binding motifs.
(E) XerD binds to XDS1 in E. coli. The indicated proteins from B. subtilis (Bs), E. coli (Ec), and S. aureus (Sa) were expressed under IPTG control. White colonies on LB(X-gal+IPTG) are indicative of protein binding to XDS1 and repression of lacZ transcription. XerD* is a catalytic XerD mutant (Y277F).
The data presented thus far are consistent with a model in which XerD bound to XDS sites functions to unload rather than stall SMC complexes. The SMC enrichment peaks at XDS sites in the terminus region were narrow (Figures 4C and S4D) and lacked a shoulder that would have resulted from a "pileup" of stalled condensin complexes. Furthermore, if XerD stalled SMC complexes at the XDS array, the /C0 122 /C14 locus harboring the array would have interacted with regions beyond its juxta-posed position at /C0 84 /C14. Similarly, SMC would have been enriched beyond the /C0 84 /C14 site (Figure 5C). We contrast the Hi-C and ChIP-seq data from strains harboring an ectopic XDS array to those from a strain in which a chromosomal locus was artificially tethered to the cytoplasmic membrane to stall SMC translocation (Figure 5D). A TetR fusion to the Tsr membrane-anchoring domain (Tsr-TetR-YFP) (Magnan et al., 2015) bound to an array of 120 tetO operators prevented DNA juxtaposition beyond the tetO array at ~122. Similarly, SMC would have been enriched beyond the ~84 site (Figure 5C). We contrast the Hi-C and ChIP-seq data from strains harboring an ectopic XDS array to those from a strain in which a chromosomal locus was artificially tethered to the cytoplasmic membrane to stall SMC translocation (Figure 5D). A TetR fusion to the Tsr membrane-anchoring domain (Tsr-TetR-YFP) (Magnan et al., 2015) bound to an array of 120 tetO operators prevented DNA juxtaposition beyond the tetO array at ~122. However, the ~122 locus interacted with a ~330 kb region on the juxtaposed DNA (Figure 5D, green arrow). Similarly, SMC was highly enriched at the tetO array and strongly depleted beyond it, while on the juxtaposed DNA there was no sharp boundary of depletion (Figure 5D, bottom). Thus, these data provide further support for the idea that XerD functions as a site-specific unloader of SMC complexes.

We took advantage of the ability to site-specifically unload condensin to investigate whether SMC complexes, once loaded, translocate autonomously and unidirectionally down the chromosome arms or whether directed movement requires a point source of newly loaded complexes. Using a strain harboring a singe origin-proximal parS site at /C0 1, (XDS)12 arrays at +26 and +19, and an IPTG-inducible allele of XerD as the sole source of the protein, we performed Hi-C on cells grown in the absence of IPTG and at 5 min intervals after its addition. Prior to IPTG addition, the interactions between the left and right arm were similar to a strain harboring a /C0 1 parS site (Figures 5E and 5G). After addition of IPTG, interactions were progressively lost starting from the terminus-proximal side of the XDS arrays (Figure 5G), while DNA juxtaposition remained unchanged between the origin-proximal parS and the arrays. Quantification of the loss of DNA juxtaposition indicates that the rate of "unzipping" (51 ± 5 kb/min) was similar to the ~50 kb/min rate of zip-up using an IPTG-inducible allele of ParB, the condensin loader (Wang et al., 2017, 2018). Thus, once loaded, SMC complexes translocate directionally and autonomously along DNA (Figure 5F).

*S. aureus* possesses the genes encoding the SMC complex, ParB, and origin-proximal parS sites (Chan et al., 2020; Kuroda et al., 2001; Livny et al., 2007) as well as xerCD and dif. To

Figure 3. Cells lacking XerD have defects in chromosome segregation
(A) 10-fold serial dilutions of the indicated strains spotted on LB agar.
(B) Representative images of the indicated strains. DAPI-stained DNA (green) and FM4-64-stained membranes (red) are shown. Scale bar indicates 4 μm. Distended nucleoids are highlighted (yellow carets). Larger fields can be found in Figure S3.
(C) Quantitative analysis of nucleoid length. Boxplot showing the mean, quartiles, minimum, and maximum of nucleoid length of the indicated strains plotted with Prism (GraphPad). Individual data points are plotted in black. Numbers are shown in Figure S2G. Two-tailed t test was performed for each pair of strains. Except for two pairs (ΔxerD versus XDSΔΔ and XDSΔΔΔΔ versus XDSΔΔΔΔxerC), all the pairs have a significant difference in the distribution of nucleoid length (p < 0.05).
(D) Merged images of replication origins (green) labeled with GFP-ParB and DAPI-stained DNA (red). Unsegregated nucleoids are highlighted (yellow carets).
investigate whether the role of XerD in SMC unloading was conserved in this bacterium, we generated a XerD-His6 fusion and performed ChIP-seq. Five XerD enrichment peaks in addition to dif were identified in the terminus region (Figure 6A). A putative XerD binding site was present within each peak that resembled the XDS sites found in B. subtilis (Figure 6B), and expression of S. aureus XerD in E. coli was sufficient to inhibit \( P^{\text{XDS}} \)-lacZ expression (Figure 2E). To establish whether XerD bound to these sites could promote condensin unloading, we introduced the XDS12 array at 40° in the S. aureus genome and analyzed the conformation of the chromosome by Hi-C. In wild-type S. aureus, the left and right chromosome arms interacted along their length, and these interactions were dependent on ParB (Figure 6C). Importantly, the XDS array blocked DNA juxtaposition beyond its insertion site at 40°. Thus, XerD/XDS-dependent unloading of SMC complexes at the replication terminus is likely to be a conserved feature of bacterial chromosome biology.

**DISCUSSION**

Altogether our data provide an integrated view of bacterial chromosome segregation in which two broadly conserved factors that play central roles in chromosome segregation at the origin and terminus have second functions in loading and unloading SMC complexes that travel between them (Figure 7). Specifically, work in several bacteria indicates that ParB proteins bound to origin-proximal \( \text{parS} \) sites are acted upon by ParA to segregate newly replicated or-chromosomes (Marbouty et al., 2015; Minnen et al., 2016; Wang et al., 2019). Origin resolution and chromosome segregation are facilitated by the action of SMC complexes that are themselves loaded at the origin by ParB/\( \text{parS} \) nucleoprotein complexes (Gruber and Errington, 2009; Sullivan et al., 2009; Wilhelm et al., 2015). Once loaded, these complexes translocate down the left and right chromosome arms in the wake of the replisomes (Marbouty et al., 2015; Minnen et al., 2016; Wang et al., 2019). During transit, the SMC complexes tether the left and right arms together, drawing sister chromatids in on themselves and away from each other (Wang et al., 2017). This directed movement likely facilitates Topoisomerase IV-mediated resolution of pre-catenanes (Burmann and Gruber, 2015) generated by the rotation of the replisome, a consequence of the torsional strain of unwinding the DNA helix (Peter et al., 1998). When the replisome reaches the terminus, dimeric and interlinked chromosomes are resolved by XerC-XerD bound to dif (Blakely et al., 1993; Sciochetti et al., 2001) and by Topoisomerase IV (Zechiedrich and Cozzarelli, 1995). Meanwhile, XerD bound to XDS sites in the terminus region unloads SMC complexes liberating them to re-load at the replication origin. The symmetry and economy in the resolution and segregation of replicating B. subtilis chromosomes described here is likely to be broadly conserved among bacteria.

We note that in strains with one or two ectopic XDS sites, SMC enrichment along the chromosome arms was largely unaffected (Figure 8D). It is unclear why the presence of one or two sites was not sufficient to unload SMC complexes and why robust un-loading required an array with 4 XDS sites. If the movement of SMC complexes involves large step sizes (Davidson et al., 2019; Ganji et al., 2018; Kim et al., 2019; Terakawa et al., 2017), then it is possible that these translocating ATPases could bypass XerD bound to a single XDS, as has been suggested for cohesin and CTCF at topologically associated domain boundaries (Hansen et al., 2017). In the case of the native XDS sites,
once *B. subtilis* condensin complexes reach the terminus with no more DNA to translocate along the stalled complexes would have multiple opportunities to encounter XerD bound to the limited number of sites in the terminus region.

The data presented here demonstrate that SMC complexes can be removed from the genome in a site-specific manner. Although we cannot rule out the possibility that XerD recruits a factor that unloads SMC, we favor a model in which XerD itself functions as the site-specific unloader. The mechanism by which XerD catalyzes SMC removal from the chromosome is currently unknown. We note that an XerD catalytic point mutant (Y277F) (Cornet et al., 1997) was able to prevent DNA juxtaposition and SMC translocation beyond an ectopic XDS array, even though it bound XDS sites less efficiently than wild-type XerD (Figures 2E, S5A, S5B, and S7). These data argue against a model in which XerD unloads condensin by cutting the chromosome and “unthreading” topologically loaded condensin rings. Instead, we favor the idea that XerD allosterically controls ring opening through its interaction with one of the proteins in the complex. A central challenge for the future is to define in molecular terms how ParB/parS loads SMC complexes onto DNA and how XerD/XDS catalyzes their removal.

**Figure 5. XerD is required for condensin unloading**

(A–C) Normalized Hi-C contact maps of strains containing a single parS at −94° (A) and arrays of 4 (B) or 12 (C) XDS sites inserted at −109°. Quantitative analysis of Hi-C data can be found in Figures S5A and S5B. Anti-SMC ChIP-seq profiles (magenta) are shown underneath the Hi-C maps. The ratio of ChIP-seq relative to −94° parS strain is plotted in blue. Black and purple arrows show the positions of parS and the XDS array. The position (−84°) juxtaposed with the −109° XDS array is shown (red arrow). SMC enrichment is reduced beyond the XDS array and the juxtaposed position, 2-fold in the strain containing XDS4 and 4-fold in XDS12. The loss of SMC enrichment from both arms is contrasted to the non-specific loss of SMC on the origin-proximal side of parS observed in Figure S1B.

(D) Hi-C and ChIP-seq plots of a strain containing a parS site at −94° parS site, (tetO)120 at −122° (green arrow), and tsr-tetR-yfp. Expression of tsr-tetR-yfp was induced (1 mM IPTG) for 1 h. Green arrow on the Hi-C map points to the interactions between the −122° locus and a ~330 kb region on the juxtaposed DNA.

(E) Hi-C contact maps of a strain containing origin at the center of the axes.

(F and G) Unzipping of juxtaposed DNA upon XerD induction in a strain with (XDS)12 arrays at −19° and +26°. Origin-centered Hi-C maps before and after IPTG addition. Time (in minutes) is shown.

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Limitations of study

The results presented in this study indicate that XerD bound to XDS sites functions in unloading SMC complexes when they reach the terminus. Although we favor the idea that XerD is itself the unloader, it is possible that XerD recruits an unknown factor to the terminus region that catalyzes SMC removal. Biochemical reconstitution using purified components will be required to unambiguously establish whether XerD possesses condensin unloading activity.

STAR METHODS

Detailed methods are provided in the online version of this paper and include the following:

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Supplemental Information

Supplemental information can be found online at https://doi.org/10.1016/j.molcel.2020.12.027.

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AUTHOR CONTRIBUTIONS

X.W. and D.Z.R. designed the study. X.W. and X.K. constructed strains and performed ChIP-seq, whole-genome sequencing, microscopy experiments, and analyses. X.W. and Z.R. performed Hi-C experiments and analyses. H.B.B. analyzed Hi-C and ChIP-seq data and performed statistical analyses. P.P. constructed strains and assisted with Hi-C experiments. X.W. and D.Z.R. wrote the manuscript.

DECLARATION OF INTERESTS

The authors declare no competing interests.
Bacterial SMC complexes are loaded onto and unloaded from chromosomes by conserved factors that function in origin segregation and terminus resolution. ParB/parS nucleoprotein complexes (light blue circles) adjacent to newly replicated origins (orange circles) are segregated toward opposite cell poles by ParB (red circles) bound non-specifically within the nucleoid. XerC and XerD (dark blue and purple circles) resolve dimeric chromosomes at dif sites (green). SMC complexes (magenta rings) are loaded at newly replicated origins and then travel down the left and right arms while tethering them together. Loop extrusion draws newly replicated sisters in on themselves and away from each other and facilitates the removal of interlinks by Topoisomerase IV (not shown). When the tethered rings reach the terminus, they are unloaded by XerD bound to XDS sites (purple).

Figure 7. Schematic model
Bacterial SMC complexes are loaded onto and unloaded from chromosomes by conserved factors that function in origin segregation and terminus resolution. ParB/parS nucleoprotein complexes (light blue circles) adjacent to newly replicated origins (orange circles) are segregated toward opposite cell poles by ParB/parS (red circles) bound non-specifically within the nucleoid. XerC and XerD (dark blue and purple circles) resolve dimeric chromosomes at dif sites (green). SMC complexes (magenta rings) are loaded at newly replicated origins by ParB/parS and then travel down the left and right arms while tethering them together. Loop extrusion draws newly replicated sisters in on themselves and away from each other and facilitates the removal of interlinks by Topoisomerase IV (not shown). When the tethered rings reach the terminus, they are unloaded by XerD bound to XDS sites (purple).


STAR METHODS

KEY RESOURCES TABLE

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RESOURCE AVAILABILITY

Lead contact
Further information and requests for resources and reagents should be directed to and will be fulfilled by the Lead Contact Xindan Wang (xindan@indiana.edu).

Materials availability
Plasmids and strains generated in this study are available from the Lead Contact with a completed Materials Transfer Agreement.

Data and code availability
Unprocessed microscopy images are available at Mendeley data: https://doi.org/10.17632/8zdhwm28nc.1. The accession number for Hi-C data (raw and analyzed) is GEO: GSE144742. The scripts for the Hi-C and statistical analyses are available from the Lead Contact upon request without restriction.

EXPERIMENTAL MODEL AND SUBJECT DETAILS

Bacillus subtilis strains and growth
Bacillus subtilis strains were derived from the prototrophic strain PY79 (Youngman et al., 1983). Cells were grown in defined rich medium (CH) (Harwood and Cutting, 1990) at 37°C with aeration. Staphylococcus aureus strains were derived from HG003 (Herbert et al., 2010) and were grown in tryptic soy broth (TSB) at 37°C with aeration. Lists of strains, plasmids, oligonucleotides and Next Generation Sequencing samples can be found in Tables S1, S2, S3, and S4.

METHOD DETAILS

Hi-C
The detailed Hi-C procedure for B. subtilis was previously described (Wang et al., 2015). Briefly, 5x10^7 cells were crosslinked with 3% formaldehyde at room temperature for 30 min then quenched with 125 mM glycine. Cells were lysed using Ready-Lyse Lysozyme (Epicenter, R1802M) followed by 0.5% SDS treatment. Solubilized chromatin was digested with HindIII for 2 hr at 37°C. The cleaved ends were filled in with Klenow and Biotin-14-dATP, dGTP, dCTP, dTTP. The products were ligated in dilute reactions with T4 DNA ligase overnight at 16°C. Crosslinks were reversed at 65°C overnight in the presence proteinase K. The DNA was then extracted twice with phenol/chloroform/isoamylalcohol (25:24:1) (PCI), precipitated with ethanol, and resuspended in 20 ml of QIAGEN EB buffer. Biotin from non-ligated ends was removed using T4 polymerase (4 hr at 20°C) followed by extraction with PCI. The DNA was then sheared by sonication for 12 min with 20% amplitude using a Qsonica Q800R2 water bath sonicator. The sheared DNA was used for library preparation with the NEBNext Ultrall kit (E7645) according to the manufacturer’s instructions for end repair, adaptor ligation, and size selection. Biotinylated DNA fragments were purified using 10 μl streptavidin beads. 5 μl DNA-bound beads were used for PCR in a 50 μl reaction for 14 cycles. PCR products were purified using Ampure beads and sequenced at the Indiana University Center for Genomics and Bioinformatics using NextSeq 550 or at the Tufts University Core facility using HiSeq 2500. Paired-end sequencing reads were mapped to the genome of B. subtilis PY79 (NCBI Reference Sequence NC_022898.1) using the same pipeline described previously (Wang et al., 2015). The B. subtilis PY79 genome was first divided into 404 10-kb bins. Subsequent analysis and visualization was done using R scripts. The genetic loci marked by degree (°) were calculated using the PY79 genome, which results in a slight shift from data published using B. subtilis 168 genomic coordinates.

Hi-C procedure for S. aureus was similar to that of B. subtilis described above, except that cells were lysed using 3 μL Ready-Lyse Lysozyme (Epicenter, R1802M) and 5 μL of 10 mg/ml lysoptaphin. Paired-end sequencing reads were mapped to the genome of S. aureus NCTC8325 genome (NCBI NC_007795.1), which was divided into 2822 10-kb bins.
ChIP-seq
Chromatin immunoprecipitation (ChIP) for B. subtilis was performed as described previously (Wang et al., 2015). Briefly, cells were crosslinked using 3% formaldehyde for 30 min at room temperature and then quenched, washed, and lysed. Chromosomal DNA was sheared to an average size of 250 bp by sonication using a Qsonica Q800R2 water bath sonicator. The lysate was then incubated overnight at 4°C with appropriate antibodies. When anti-SMC (Lindow et al., 2002) or anti-GFP (Rudner et al., 1999) antibodies were used, lysates were incubated with Protein A Sepharose (GE Healthcare) for 1h at 4°C. When Protein C antibodies (Zheng et al., 2013) were used, lysates were included with Protein G Sepharose (GE Healthcare). After washes and elution, the immunoprecipitate was incubated at 65°C overnight to reverse the crosslinks. The DNA was further treated with RNaseA, Proteinase K, extracted with PCI, resuspended in 50 μl EB and used for library preparation with the NEBNext Ultra II kit (E7645) and sequenced using the Illumina MiSeq or NextSeq550 platforms. The sequencing reads were aligned to the B. subtilis PY79 genome (NCBI NC_022898.1) using CLC Genomics Workbench (CLC Bio, QIAGEN), and subsequently normalized, plotted and analyzed using R scripts.

ChIP for S. aureus was performed similar B. subtilis, except that 1) cells were lysed using 3 μl Ready-Lyse Lysozyme (Epicenter, R1802M) and 80 μL of 10 mg/ml lysostaphin; 2) anti-His antibodies (Genescript 10498-106) and Protein G-Sepharose (GE Healthcare) were used; and 3) sequencing reads were mapped to the genome of S. aureus NCTC8325 (NCBI NC_007795.1).

ChIP-seq analysis
Sequencing reads from ChIP and input samples were normalized to the total number of reads. The ChIP enrichment (ChIP/Input) was plotted. After excluding enriched regions containing highly transcribed genes which are enriched in every ChIP-seq experiment we have done regardless of the strain or antibodies used (Wang et al., 2017), we manually identified peaks with greater than 3-fold ChIP/input enrichment. For XerC-GFP (Figure 2A) and XerC-PrC (Figure S2A), we identified a single peak at dif, which was statistically significant in both samples (Figure S2E). For two replicates of XerD-GFP (Figures 2B and 2C) and one sample of XerD-PrC (Figure S2B), in addition to the dif site, we identified four sites (XDS1-4) statistically significant in all three samples and a fifth site (XDS5) significant in two out of the three samples (Figure S2E). Details on the statistical tests can be found in the legend of Figure S2E.

For Hi-C experiments, two biological replicates were performed in strains containing the XDS array was inserted at different genomic positions. All of these experiments were intermixed and did not allow for a traditional quantitative analysis. Figures in which different ectopic parS sites were analyzed by Hi-C include Figures 1, 5, S1, S5, and S6. Figures in which different ectopic XDS sites were analyzed by Hi-C include Figures 5, S5, and S6.

Biological replicates
For the ChIP-seq analysis of XerD binding sites, two biological replicates were performed using XerD-GFP and one replicate using XerD-PrC. Control ChIP-seq experiments with XerC-GFP and XerC-PrC were each performed one time. These replicates are included in Figures 2B, 2C, S2A, and S2B with quantitative analysis in Figure S2E.

For the ChIP-Seq analysis of SMC enrichment along the chromosome arms and the specific enrichment at XDS sites, two biological replicates were performed in a strain in which parS was inserted at −117° and two replicates in a strain in which parS was inserted at −94°. These replicates are included in Figure S4A.

For Hi-C experiments, two biological replicates were performed in strains containing −117° parS in the presence and absence of xerD. These can be found Figure S1D. The replicates were highly reproducible. Most of the other strains analyzed in this study by Hi-C were performed only once. However, the conclusions were drawn from Hi-C data obtained using complementary strains. For example, the effect of XDS sites on DNA juxtaposition was analyzed in strains in which parS was present at several distinct locations in the genome and in strains in which the XDS array was inserted at different genomic positions. All of these experiments were internally consistent providing support for our conclusions but do not allow for a traditional quantitative analysis. Figures in which different ectopic parS sites were analyzed by Hi-C include Figures 1, 5, S1, SS, and S6. Figures in which different ectopic XDS sites were analyzed by Hi-C include Figures 5 and S5.

Microscopy
Fluorescence microscopy was performed on a Nikon Ti2E microscope equipped with Plan Apo 100x/1.4NA phase contrast oil objective and an sCMOS camera. Cells were immobilized using 2% agarose pads containing growth media. Membranes were stained with FM4-64 (Molecular Probes) at 3 μg/ml. DNA was stained with DAPI at 2 μg/ml. Images were cropped and adjusted using MetaMorph software (Molecular Devices). Final figures were prepared in Adobe Illustrator.

Plasmid construction
pWX118L [yvrN (−122):tetO120 (cat)] was generated by inserting tetO120 (liberated with Nhel and HindIII from pLAU39 CGSC#: 12311 (Lau et al., 2003)) into pWX109 between Nhel and HindIII. pWX109 [yvrN::cat] is an ectopic integration vector for double crossover insertions into the yvrN locus (X.W. and D.Z.R., unpublished data). The genetic location by degrees (°) in this study is calculated using the PY79 genome sequence (CP006881) (Schroeder and Simmons, 2013), which is slightly different from our previous publications using B. subtilis 168 genome coordinates (NC_000964) (Kunst et al., 1997).
pWX618 [peB::Psoj (optRBS) mgfpmut3-spo0J (ΔparS) (cat)] was constructed in a 3-way ligation to insert mgfpmut3 with an optimal ribosome binding site (optRBS) (amplified using oWX706 and oWX674 from pDHL580 (Landgraf et al., 2012) and digested with HindIII and Xhol) and spo0J (ΔparS) (excised from pKM256 using Xhol and BamHI) into pKM170 [peB::Psoj (cat)] (Sullivan et al., 2009) between HindIII and BamHI. spo0J (ΔparS) has 7 synonymous changes in the 16-base parS site in the spo0J (Sullivan et al., 2009). parB was named spo0J in B. subtilis.

pWX702 [yvbJ::Pspank (optRBS) tsr-tetr-yfp (erm)] was generated in a 3-way ligation to insert tsr (Magnan et al., 2019) with an optimal ribosome binding site (optRBS) (amplified using oWX1429 and oWX1430 from E. coli MG1655 genome and digested with HindIII and XbaI) and tetr-yfp (amplified using oWX1431 and oWX1432 from pWX187 and digested with XbaI and Xmal) into pMS44 [yvbJ::Pspank (erm)] (D.Z.R., unpublished data) between HindIII and Xmal. The construct was sequenced using oWX486, oWX487, oWX669, oWX1434, and oWX1435.

pWX720 [yvbJ::Pspank (optRBS) xerD (cat)] was generated by inserting xerD with an optimal ribosome binding site (optRBS) (amplified using oWX1645 and oWX1646 from B. subtilis PY79 genome and digested with Xmal and Nhel) into pER134 [yvbJ::Pspank (cat)] (D.Z.R., unpublished data) between Xmal and Nhel. The construct was sequenced using oWX486 and odr829.

pWX725 [Ptet (optRBS) Sa xerD-his6 (cat)] was generated by inserting Sa xerD with an optimal ribosome binding site (optRBS) (amplified using oWX1706 and oWX1707 from S. aureus HG003 (Herbert et al., 2010) genome and digested with KpnI and Xmal) into pTP171 (Pang et al., 2017) between KpnI and Xmal. pTP171 is an S. aureus integration plasmid that contains Ptet (optRBS) Bs noc-his6 cat. The construct was sequenced using oWX1710 and oWX1711.

pWX726 (XDS12 (loxP-kan-loxP]) was generated by an isothermal assembly reaction containing 2 PCR products: 1) (XDS12 IoxP- kan-loxP] (amplified using oWX1563 and oWX438 from the genomic DNA of BWX3805 (see strain construction); 2) pTP171 (Pang et al., 2017) amplified using oWX1712 and oWX1713. The construct was sequenced using oTP263 and oMLB8.

pWX730 [attB::cat Plac::Bs xerD] was generated by an isothermal assembly reaction containing 2 PCR products: 1) Bs xerD (amplified using oWX1721 and oWX1722 from PY79 genomic DNA; 2) pHC886 [attB::cat Plac empty vector] (H. Cho and T. G. Bernhardt, personal communication) amplified using oWX1397 and oWX1398. The construct was sequenced using oWX1403 and oWX1404.

pWX731 [attB::cat Plac::Bs xerD (Y277F)] was generated by an isothermal assembly reaction containing 2 PCR products: 1) Bs xerD Y277F (amplified using oWX1721 and oWX1722 from genomic DNA of BWX3896 (see strain construction); 2) pHC886 [attB::cat Plac empty vector] (H. Cho and T. G. Bernhardt, personal communication) amplified using oWX1397 and oWX1398. The construct was sequenced using oWX1403 and oWX1404.

pWX732 [attB::cat Plac::Bs xerC] was generated by an isothermal assembly reaction containing 2 PCR products: 1) Bs xerC (amplified using oWX1723 and oWX1724 from PY79 genomic DNA; 2) pHC886 [attB::cat Plac empty vector] (H. Cho and T. G. Bernhardt, personal communication) amplified using oWX1397 and oWX1398. The construct was sequenced using oWX1403 and oWX1404.

pWX737 [attB::cat Plac::Ec xerD] was generated by an isothermal assembly reaction containing 2 PCR products: 1) Ec xerD (amplified using oWX1725 and oWX1726 from E. coli MG1655 genomic DNA; 2) pHC886 [attB::cat Plac empty vector] (H. Cho and T. G. Bernhardt, personal communication) amplified using oWX1397 and oWX1398. The construct was sequenced using oWX1403 and oWX1404.

pWX738 [attB::cat Plac::Sa xerD] was generated by an isothermal assembly reaction containing 2 PCR products: 1) Sa xerD (amplified using oWX1727 and oWX1728 from E. coli MG1655 genomic DNA; 2) pHC886 [attB::cat Plac empty vector] (H. Cho and T. G. Bernhardt, personal communication) amplified using oWX1397 and oWX1398. The construct was sequenced using oWX1403 and oWX1404.

Strain construction (B. subtilis)

The `rtp`, `spollIE`, `sftA`, `xerC` and `xerD` mutants marked with loxP-erm-loxP are derived from the Bacillus subtilis knockout collection (Koo et al., 2017). Genomic DNA of these knockout strains was extracted and backcrossed with PY79 twice. Each deletion was confirmed by PCR using gKO0 (within the erythromycin resistance cassette) and an upstream gene-specific primer: for rtp (oWX1186), xerC (oWX1448), xerD (oWX1187), spollIE (oWX1175), and sftA (oWX1176). In some cases, the loxP-erm-loxP cassette was removed using a cre expressing plasmid pDR244 (Meeseke et al., 2015).

+diff loxP-kan-loxP (BWX3510) or loxP-spec-loxP (BWX3508).

The dif site (ACTTCTCTAGAATATATAGTGAACAT) was deleted by directly transforming an isothermal assembly product to PY79. The isothermal assembly reaction contained 3 PCR products: 1) the region upstream of dif (amplified from PY79 genomic DNA using oWX1377 and oWX1378); 2) loxP-kan-loxP cassette or loxP-spec-loxP cassette (amplified from pWX470 or pWX466 using universal primers oWX438 and oWX439) and 3) the region downstream of dif (amplified from PY79 genomic DNA using primers oWX1379 and oWX1380). The transforms were amplified and sequenced using oWX1381 and oWX1382.

+58 parS loxP-kan-loxP (BWX3388).

The +4 `parS` sequence (TGTTACAGTGAACAT) was inserted at +58 (in the intergenic region between yeaB and yeaC). An isothermal assembly product was directly transformed to parS9 (BWX3212) (Wang et al., 2015), which has all the 9 `parS` sites deleted from the B. subtilis genome. The isothermal assembly reaction contained 3 PCR products: 1) a region containing yeaB (amplified from PY79 genomic DNA using oWX1309 and oWX1310); 2) loxP-kan-loxP cassette flanked by the +4 `parS` sequence (amplified from pWX470 using universal primers oWX1241 and oWX438) and 3) a region containing yeaC (amplified from PY79 genomic DNA using primers oWX1311 and oWX1312). The transforms were amplified and sequenced using oWX1313 and oWX1314.
+117 parS loxP-kan-loxP (BWX3389).
The +4° parS sequence (TGTTACGTGAAACA) was inserted at +117° (in the intergenic region between ykcC and htra). An isothermal assembly product was directly transformed to parS J9 (BWX3212) (Wang et al., 2015). The isothermal assembly reaction contained 3 PCR products: 1) a region containing ykcC gene (amplified from PY79 genomic DNA using oWX1315 and oWX1316); 2) loxP-kan-loxP cassette flanked by the +4° parS sequence (amplified from pWX470 using universal primers oWX1241 and oWX438) and 3) a region containing htra (amplified from PY79 genomic DNA using primers oWX1317 and oWX1318). The transformants were amplified and sequenced using oWX1319 and oWX1320.

+153 parS loxP-kan-loxP (BWX3391).
The +4° parS sequence (TGTTACGTGAAACA) was inserted at +153° (in the intergenic region between ymfJ and ymfK). An isothermal assembly product was directly transformed to parS J9 (BWX3212) (Wang et al., 2015). The isothermal assembly reaction contained 3 PCR products: 1) a region containing ymfJ gene (amplified from PY79 genomic DNA using oWX1321 and oWX1322); 2) loxP-kan-loxP cassette flanked by the +4° parS sequence (amplified from pWX470 using universal primers oWX1241 and oWX438) and 3) a region containing ymfK (amplified from PY79 genomic DNA using primers oWX1323 and oWX1324). The transformants were amplified and sequenced using oWX1325 and oWX1326.

recA loxP-spec-loxP (BWX3649).
The recA gene was deleted by directly transforming an isothermal assembly product to PY79. The isothermal assembly reaction contained 3 PCR products: 1) the region upstream of recA (amplified from PY79 genomic DNA using oWX1441 and oWX1442); 2) loxP-spec-loxP cassette (amplified from pWX466 using universal primers oWX438 and oWX439) and 3) the region downstream of recA (amplified from PY79 genomic DNA using primers oWX1443 and oWX1444). The transformants were amplified and sequenced using oWX1445 and oWX1446.

xerC-mgfpmut3 spec (BWX3653).
An isothermal assembly reaction containing the following 3 PCR products was directly transformed to PY79: 1) a region containing xerC and upstream region (amplified from PY79 genomic DNA using oWX1447 and oWX1448); 2) mgfpmut3 spec (amplified from pWX429 using oWX1385 and oWX1386); 3) a region downstream of xerC (amplified from PY79 genomic DNA using primers oWX1450 and oWX1451). The transformants were amplified using oWX1452 and oWX1453 and sequenced using oWX1452 and oML79. The transformants contain ycgO-PftsW lacI-mgfpmut3 spec (X.W. and D.Z.R., unpublished data).

xerD-mgfpmut3 spec (BWX3655).
An isothermal assembly reaction containing the following 3 PCR products was directly transformed to PY79: 1) a region containing xerD and upstream region (amplified from PY79 genomic DNA using oWX1454 and oWX1455); 2) mgfpmut3 spec (amplified from pWX429 using oWX1385 and oWX1386); 3) the region downstream of xerD (amplified from PY79 genomic DNA using primers oWX1456 and oWX1457). The transformants were amplified using oWX1458 and oWX1459 and sequenced using oWX1458 and oML79.

yrdq (2540kb)::XDS1 loxP-kan-loxP (BWX3724).
The XDS1 site at 1717 kb was inserted between yrdQ and yrdP by transforming an isothermal assembly product to PY79. The isothermal assembly reaction contained 4 PCR products: 1) a region containing yrdQ (amplified from PY79 genomic DNA using oWX1504 and oWX1505); 2) XDS1 at 1717 kb (amplified from the PY79 genomic DNA using oWX1502 and oWX1503) 3) loxP-kan-loxP cassette (amplified from pWX470 using universal primers oWX438 and oWX439) and 4) a region containing yrdP (amplified from PY79 genomic DNA using primers oWX1506 and oWX1507). The transformants were amplified using oWX1508 and oWX1509, and sequenced using oWX1509.

-109° (XDS)4 loxP-spec-loxP (BWX3790).
The (XDS)4 array was synthesized at Integrated DNA Technologies (IDT) as gBlockWX06, which contains 50 bp regions centered at XDS1 (1717kb), XDS2 (1912kb), XDS3 (1989kb), XDS4 (2033kb), and two 20 bp sequences at the 5’ and 3’ end of the construct for primer binding. This array was inserted at −109° (2802 kb) using an isothermal assembly reaction that contained 4 PCR products: 1) the region upstream of 2802 kb (amplified from PY79 genomic DNA using oWX1551 and oWX1552); 2) gBlockWX06; 3) loxP-spec-loxP cassette (amplified from pWX466 using universal primers oWX438 and oWX439); 4) the region downstream of 2802 kb (amplified from PY79 genomic DNA using primers oWX1553 and oWX1554). The transformants were amplified using oWX1555 and oWX1556, and sequenced using oWX1555.

-109° (XDS)12 loxP-kan-loxP (BWX3805).
The (XDS)12 array was synthesized at Integrated DNA Technologies (IDT) as gBlockWX07, which contains 30 bp regions centered at XDS1 (1717kb), XDS2 (1912kb), XDS3 (1989kb) and XDS4 (2033kb) repeated 3 times, and 20 bp unique spacer sequences between the XDS sites to reduce recombination. This fragment also contains two 20 bp sequences at the 5’ end and 3’ end for primer binding. This array was inserted at −109° (2802 kb) using an isothermal assembly reaction that contained 4 PCR products: 1) the region upstream of 2802 kb (amplified from PY79 genomic DNA using oWX1551 and oWX1552); 2) gBlockWX07; 3) loxP-kan-loxP cassette (amplified from pWX470 using universal primers oWX438 and oWX439); 4) the region downstream of 2802 kb (amplified from PY79 genomic DNA using primers oWX1553 and oWX1554). The transformants were amplified using oWX1555 and oWX1556, and sequenced using oWX1555. In some cases, the loxP-kan-loxP cassette was removed using a cre expressing plasmid pDR244 (Meeske et al., 2015).
The (XDS)12 array was inserted at –80° (3128 kb) using an isothermal assembly reaction that contained 4 PCR products: 1) the region upstream of 3128 kb (amplified from PY79 genomic DNA using oWX1551 and oWX1552); 2) xerD-prC spec (BWX3970). The Y277F point mutation was generated by transforming an isotheral assembly reaction containing the following 2 PCR products directly transformed to PY79: 1) xerD and its upstream region (amplified from PY79 genomic DNA using oWX1454 and oWX1618); 2) xerD-prC spec (BWX3970). An isothermal assembly reaction containing the following 2 PCR products was directly transformed to PY79: 1) region containing xerC and upstream region (amplified from PY79 genomic DNA using oWX1448 and oWX1644); 2) region containing spec and the downstream of xerD (amplified from PY79 genomic DNA using primers oWX1643 and oWX1457). The transformants were amplified using oWX1500 and oWX1501, and sequenced using oWX1500 and oML77.

**xerD-prC spec (BWX3974)**

An isothermal assembly reaction containing the following 2 PCR products was directly transformed to PY79: 1) region containing xerC and upstream region (amplified from PY79 genomic DNA using oWX1448 and oWX1644); 2) region containing spec and the downstream of xerD (amplified from PY79 genomic DNA using primers oWX1643 and oWX1451). This results in the addition of a 5 amino acid linker (LEGSG) and a PrC tag (EDQVDPRLIDGK) to the C terminus of XerD before the stop codon. The transformants were amplified using oWX1500 and oWX1501, and sequenced using oWX1500 and oML79.

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**loxP-spec-loxP (BWX3855)**

This strain was constructed using the same method as BWX3805 except that a loxP-spec-loxP cassette was used (amplified from pWX466 using universal primers oWX438 and oWX439).

**loxP-spec-loxP (BWX3858)**

The (XDS)12 array was inserted at –80° (3128 kb) using an isothermal assembly reaction that contained 4 PCR products: 1) the region upstream of 3128 kb (amplified from PY79 genomic DNA using oWX1551 and oWX1552); 2) xerD-prC spec (BWX3970). The Y277F point mutation was generated by transforming an isotheral assembly reaction containing the following 2 PCR products directly transformed to PY79: 1) xerD and its upstream region (amplified from PY79 genomic DNA using oWX1454 and oWX1618); 2) xerD-prC spec (BWX3970). An isothermal assembly reaction containing the following 2 PCR products was directly transformed to PY79: 1) region containing xerC and upstream region (amplified from PY79 genomic DNA using oWX1448 and oWX1644); 2) region containing spec and the downstream of xerD (amplified from PY79 genomic DNA using primers oWX1643 and oWX1451). This results in the addition of a 5 amino acid linker (LEGSG) and a PrC tag (EDQVDPRLIDGK) to the C terminus of XerD before the stop codon. The transformants were amplified using oWX1500 and oWX1501, and sequenced using oWX1500 and oML79.

**loxP-spec-loxP (BWX4004)**

The (XDS)12 loxP-spec-loxP fragment was inserted at +26° in BWX3370 (Wang et al., 2017) that contains a single parS site at –1° using an isothermal assembly reaction that contained 3 PCR products: 1) the region upstream of +26° (amplified from PY79 genomic DNA using oWX1336 and oWX1654); 2) (XDS)12 loxP-spec-loxP fragment amplified from the genomic DNA of BWX3970 using universal primers oWX438 and oWX1563; 3) the region downstream of +26° (amplified from PY79 genomic DNA using primers oWX1338 and oWX1339). The transformants were amplified using oWX1340 and oWX1341, and sequenced using oWX1340 and oML83. In some cases, the loxP-spec-loxP cassette was removed using a cre expressing plasmid pDR244 (Meeske et al., 2015).

**loxP-spec-loxP (BWX4006)**

The (XDS)12 loxP-spec-loxP fragment was inserted at –19° using an isothermal assembly reaction that contained 3 PCR products: 1) the region upstream of –19° (amplified from PY79 genomic DNA using oWX1648 and oWX1649); 2) (XDS)12 loxP-spec-loxP fragment amplified from the genomic DNA of BWX3970 using universal primers oWX438 and oWX1563; 3) the region downstream of –19° (amplified from PY79 genomic DNA using primers oWX1650 and oWX1651). The transformants were amplified using oWX1652 and oWX1653, and sequenced using oWX1652 and oML79. In some cases, the loxP-spec-loxP cassette was removed using a cre expressing plasmid pWX492 (X.W. and D.Z.R., unpublished data).
-109 (XDS)1 loxP-spec-loxP (BWX4201).

An isothermal assembly reaction containing the following 2 PCR products was directly transformed to PY79: 1) a region containing the upstream of 2802 kb region (amplified from BWX3855 genomic DNA using oWX1551 and oWX1729); 2) a region containing loxP-spec-loxP and the downstream of 2802 kb region (amplified from BWX3855 genomic DNA using primers oWX439 and oWX1554). This results in retaining a single XDS site from the (XDS)12 array. The transformants were amplified using oWX1555 and oWX1556 and sequenced using oWX1555.

-109 (XDS)2 loxP-spec-loxP (BWX4203).

An isothermal assembly reaction containing the following 2 PCR products was directly transformed to PY79: 1) a region containing the upstream of 2802 kb region (amplified from BWX3855 genomic DNA using oWX1551 and oWX1730); 2) a region containing loxP-spec-loxP and the downstream of 2802 kb region (amplified from BWX3855 genomic DNA using primers oWX1731 and oWX1554). This results in retaining two XDS sites from the (XDS)12 array. The transformants were amplified using oWX1555 and oWX1556 and sequenced using oWX1555.

\section*{Strain construction (E. coli)}

\underline{FTR-kan-FRT} PXDS1-lacZ in E. coli.

To construct the lacZ reporter repressed by XerD, we used a method described in (Cho and Bernhardt, 2013). Specifically, the chromosomal region of TB10 (Johnson et al., 2004) encompassing the lacI gene and lac promoter was replaced with a kanamycin resistance cassette (FTR-kan-FRT) and a synthetic promoter containing the XerD binding sequence at 1717 kb (XDS1) inserted between the -35 and -10 elements of the promoter by λ Red recombineering. An isothermal assembly reaction that contained the following 2 PCR products was transformed to TB10 by electroporation: 1) a region upstream of lacZ (amplified from E. coli strain HC328 (Cho and Bernhardt, 2013) using oWX1717 and oWX1718); 2) a region containing lacZ (amplified from HC328 using oWX1719 and oWX1720). The resulting E. coli construct was amplified using oWX1717 and oWX1720 and sequenced using oWX1720. Following recombineering, the FTR-kan-FRT PXDS1-lacZ reporter was transduced into MG1655 using P1 phage, resulting cWX1495.

\underline{att} Plac expressing constructs.

pHC886, pWX730, pWX731, pWX732, pWX737 and pWX738 were integrated to attP using methods described in (Haldimann and Wanner, 2001). Specifically, the plasmids was transformed into TB28 (Bernhardt and de Boer, 2004) containing pNT-ts (Haldimann and Wanner, 2001). The integration was confirmed by amplification using oligos attP1, attP4, CRIM P2, CRIM P3 (Haldimann and Wanner, 2001). The constructs were transduced into oWX1495 by P1 phage.
Strain construction (S. aureus)

**geh(40/C14)::pWX725 (Ptet Sa xerD-his6) (SaWX003)**

This strain was constructed by electroporating plasmid pWX725 into strain aTP176 (RN4220, pTP044) (Pang et al., 2017) and selected on TSB plate containing chloramphenicol (5 μg/ml). In the presence L54a integrase expressed from pTP044 (Pang et al., 2017), pWX725 integrates into the attB(L54a) site within the geh gene of S. aureus, at 40° of the genome. The integration was confirmed using oTP133-136 and resulting strain was amplified sequenced using oWX1710 and 1711. To perform ChiP-seq, this construct was transduced into aTP394 (Pang et al., 2017) which has a transposon insertion in the spa gene encoding Surface Protein A in HG003 (Herbert et al., 2010), resulting in SaWX019. The transducing phage used was phage φ11.

**geh(40/C14)::pWX726 ((XDS)12 loxP-kan-loxP) (SaWX005)**

This strain was constructed in a way similar to SaWX003. Specifically, plasmid pWX726 was electroporated into strain aTP176 (RN4220 + pTP044) to integrate into geh and selected on TSB plate containing kanamycin (25 μg/ml) and neomycin (25 μg/ml). The integration was confirmed using oTP133-136 and resulting strain was amplified sequenced using oTP263 and oML83. To perform Hi-C, this construct was transduced into HG003 (Herbert et al., 2010), by using phage φ11. The resulting strain is SaWX013.

**QUANTIFICATION AND STATISTICAL ANALYSIS**

**Analysis of nucleoid length**

The length of DAPI-stained nucleoid was measured using MetaMorph (Molecular Devices) and presented in Figure S2G. N represents the number of nucleoids. Mean and standard deviation (std dev) are shown. The distribution of the data is plotted in Figure 3C using Prism (GraphPad). Two-tailed t test was performed for each pair of strains using Prism.

**Analysis of XerC and XerD ChiP-Seq peaks**

ChiP-seq peaks for XerC and XerD were identified as described in METHOD DETAILS. In Figure S2E, we performed a statistical test using the Model-based Analysis of ChiP-Seq (MACS) procedure (Zhang et al., 2008) using custom scripts written in Python 3.7.6 (https://www.python.org/). The p values were computed with the scipy.stats.poisson.cdf function from the Scipy 1.4.1 module (Virtanen et al., 2020). The procedure is described in detail in the figure legend.

**Analysis of SMC ChiP-Seq peaks at XDS sites**

For statistical test for SMC enrichment peaks at XDS sites in Figure S4C, p values were computed using a Mann-Whitney U test with the scipy.stats.mannwhitneyu function of the Scipy 1.4.1 module (Virtanen et al., 2020) in Python 3.7.6 (https://www.python.org/). Details of the analysis can be found in the figure legend.